

Synthesis of L-Enantiomers of 4'-Thioarabinofuranosyl Pyrimidine Nucleosides

Hiroshi Satoh, Yuichi Yoshimura,* Shinji Sakata, Shinji Miura, Haruhiko Machida

*Biochemicals Division, Yamasa Corporation,
2-10-1 Araocho, Choshi, Chiba 288, Japan*

Akira Matsuda

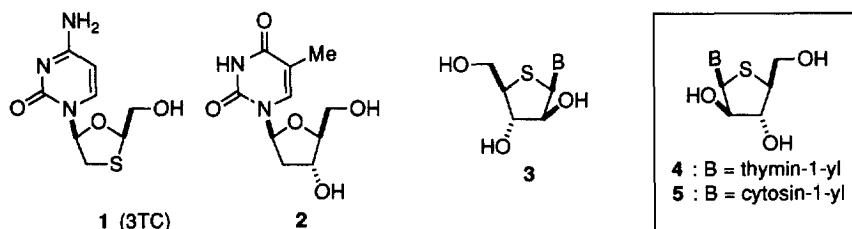
*Faculty of Pharmaceutical Sciences, Hokkaido University;
Kita-12, Nishi-6, Kita-ku, Sapporo 060, Japan*

Received 18 February 1998; accepted 18 March 1998

Abstract: L-Enantiomers of 4'-thioarabinofuranosyl pyrimidine nucleosides were synthesized from D-xylose. Methyl 2,3,5-tri-O-benzyl-D-xylofuranoside **6** was converted to the corresponding xylitol **7**, which was treated with MsCl and then Na₂S to give 1,4-anhydro-L-4-thioarabitol **8**. As previously reported, Pummerer rearrangement of **8** followed by glycosylation with a silylated thymine and N4-acetylcytosine derivative and deprotection gave the corresponding α - and β -L-4'-thioarabinofuranosyl pyrimidine nucleosides.

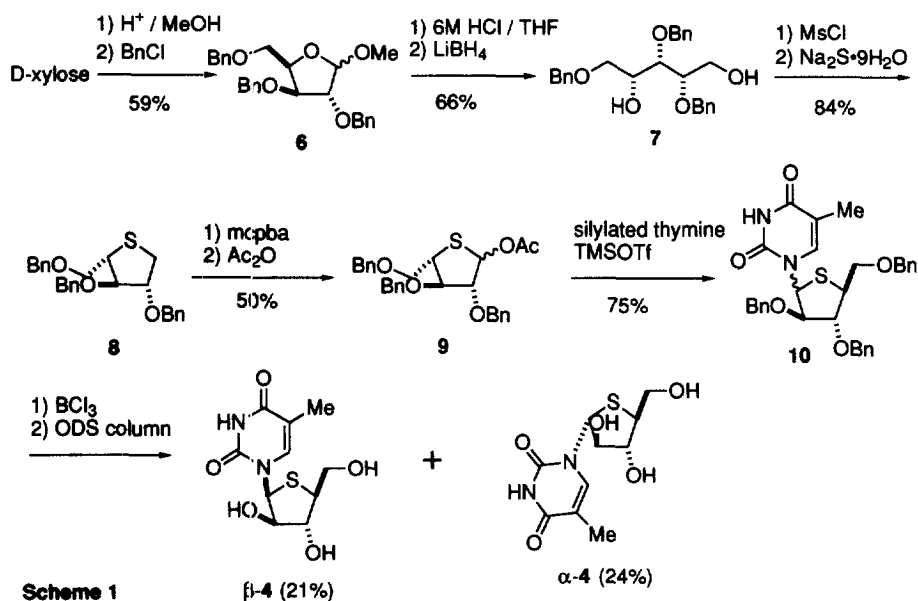
© 1998 Elsevier Science Ltd. All rights reserved.

Nucleoside antimetabolites are known to inhibit the synthesis of DNA or RNA after conversion to the corresponding triphosphate analogues, and thus have antiviral and antineoplastic activities.¹ Until 1992, the biologically active analogues, which should be phosphorylated by kinases, were believed to be the D-enantiomers, as with naturally occurring nucleosides. However, this preconception changed with the discovery of the potent anti-human immunodeficiency virus type 1 activity of L-(-)-3TC **1**, which exhibited less cytotoxicity than its D-enantiomer.² It was also reported that L-thymidine **2** was a substrate of thymidine kinase that was coded by herpes viruses and had weak anti-herpes simplex virus type 1 (HSV-1) activity.³ Therefore, L-nucleosides have been considered as potential selective antiviral agents.⁴



We focused on the L-enantiomers of 4'-thioarabinonucleosides as potential antiviral agents. We recently exploited a facile synthesis of D-4'-thioarabinonucleosides **3** and reported their potent antiherpes viral

activities.⁵ Although many other syntheses of D-4'-thionucleosides have been reported,⁶ the synthesis of their L-enantiomers has been limited.⁷ To the best of our knowledge, there has been no report concerning the synthesis of L-4'-thioarabinonucleosides. Thus, we describe here a novel and convenient synthesis of L-4'-thioarabinonucleosides from D-xylose.



Methyl 2,3,5-tri-*O*-benzyl-D-xylofuranoside **6**, which was easily obtained from D-xylose in 2 steps, was hydrolyzed under acidic conditions and reduced by LiBH₄ in THF to give xylitol **7** in 66% yield from **6**. As originally reported in the synthesis of 4'-thioDMDC and 4'-thiogemcitabine,⁸ xylitol **7** was converted to its dimesylate, which was treated with sodium sulfide in DMF at 100 °C for 3 h to give 1,4-anhydro-4-thio-L-arabitol **8** in 84% yield. Following the synthesis of D-4'-thioarabinonucleosides,⁵ oxidation of **8** with *m*-chloroperbenzoic acid (mcpba) at -78 °C in CH₂Cl₂ gave a diastereo mixture of the sulfoxides, which was subjected to Pummerer rearrangement with acetic anhydride to give an anomeric mixture of L-4-thioarabinose **9** in 50% yield from **8** (Scheme 1).

Table 1: Optical Rotations of D- and L-Enantiomers of **8** and **9**^a

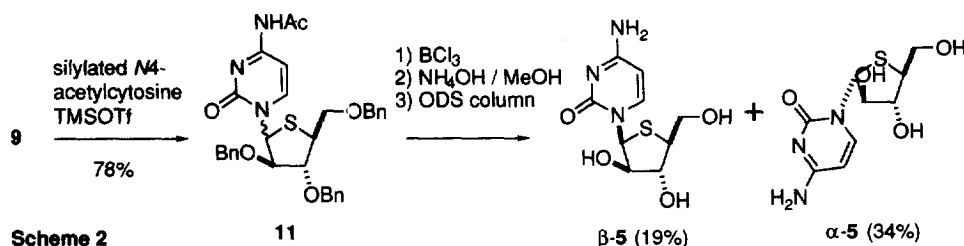
	D- 8 ^b	L- 8	D- 9 ^b	L- 9
[α] _D ²⁵	+0.32° (<i>c</i> = 2.5)	-0.38° (<i>c</i> = 2.5)	-30.6° (<i>c</i> = 2.0)	+29.8° (<i>c</i> = 2.0)

^aAll of the optical rotations were measured in CHCl₃ at 25 °C. ^bYoshimura, unpublished data

To confirm the stereochemistry of the resulting compounds, the optical rotations of **8** and **9** were compared with those of their D-enantiomers, which were synthesized previously.⁵ Although 4-thioarabinose **9** was an anomeric mixture, separation of the mixture was unsuccessful. However, the ratio of the anomers was identical for the D- and L-enantiomers (α : β = 1 : 2), thus, the optical rotations of D- and L-**9** were measured as

they were. The results, summarized in Table 1, clearly supported the L-stereochemistry of **8** and **9**, as we expected. Previous synthesis of the D-enantiomers **8** and **9** was also started from a D-xylofuranoside derivative.⁵ It is noteworthy that both D- and L-4-thioarabinose moiety could be synthesized from the same starting material by shifting the corresponding chiral carbons to use.

The glycosylation reaction between L-4-thioarabinose **9** and a persilylated thymine derivative in the presence of trimethylsilyl trifluoromethanesulfonate (TMSOTf) gave an anomeric mixture of benzylated L-4'-thioarabinosylthymine **10** in 75% yield. Debenzoylation of **10** by treatment with BCl₃ in CH₂Cl₂ at -20 °C, followed by separation of the anomers using ODS reversed-phase column chromatography gave α - and β -4'-thioarabinosylthymine **4** in yields of 24% and 21%, respectively.⁹ The determination of the α - and β stereochemistry was made by ¹H NMR and other instrumental analyses in comparison with α - and β -D-**4**. In a similar manner, glycosylation using persilylated N4-acetylcytosine with **9** gave 4'-thionucleoside **11**, which was deprotected (2 steps) to give α - and β -L-4'-thioarabinosylcytosine **5**¹⁰ (Scheme 2).



The antiviral activities of α -**4**, β -**4**, and β -**5** were evaluated against HSV-1 and herpes simplex virus type 2 (HSV-2). However, they did not show any activities up to 100 μ g/mL. These compounds were not cytotoxic against human T-cell leukemia, CCRF-HSB-2, up to 100 μ g/mL. Only α -**5** showed moderate anti-HSV-1 activity (ED₅₀ = 10 μ g/mL) without cytotoxicity against CCRF-HSB-2 (IC₅₀ >100 μ g/mL). α -**5** was also active against HSV-2 (ED₅₀ = 10 μ g/mL). Only the α -L-cytidine derivative, and not the β -L-cytidine derivative, showed antiviral activity. These results are contrasted with those of D-4'-thioarabinonucleosides: β -D-isomers possess potent anti-herpesvirus activities, while α -D-isomers are inactive.⁵

In summary, we developed a novel and convenient synthesis of L-enantiomers of 4'-thioarabinofuranosyl pyrimidine nucleosides from D-xylose. Further synthesis of L-4'-thioarabinonucleoside derivatives are underway.

Acknowledgements. The authors would like to thank Dr. K. Kodama for his encouragement throughout this work. The authors also thank Mr. M. Morozumi for his helpful discussions.

References and Notes

- For example, see: Ono, T.; Fujii, A.; Yamagami, K.; Hosoya, M.; Okumoto, T.; Sakata, S.; Matsuda, A.; Sasaki, T. *Biochem. Pharmacol.* **1996**, *52*, 1279–1285.
- a) Coates, J. A. V.; Cammack, N.; Jenkinson, H. J.; Mutton, I. M.; Pearson, B. A.; Storer, R.; Cameron, J. M.; Penn, C. R. *Antimicrob Agents Chemother.* **1992**, *36*, 202–205. b) Schinazi, R. F.; Chu, C. K.; Peck, A.; McMillan, A.; Mathis, R.; Cannon, D.; Jeong, L.-S.; Beach, J. W.; Choi, W.-B.; Yeola, S.; Liotta, D. C. *Antimicrob. Agents Chemother.* **1992**, *36*, 672–676. c) Coates, J. A. V.; Cammack, N.; Jenkinson, H.

- J.; Jowett, A. J.; Jowett, M. I.; Pearson, B. A.; Penn, C. R.; Rouse, P. L.; Viner, K. C.; Cameron, J. M. *Antimicrob. Agents Chemother.* **1992**, *36*, 733–739.
3. Spadari, S.; Maga, G.; Focher, F.; Ciarrochi, G.; Manservigi, R.; Arcamone, F.; Capobianco, M.; Carcuro, A.; Colonna, F.; Iotti, S.; Garbesi, A. *J. Med. Chem.* **1992**, *35*, 4214–4220.
 4. Kotra, L. P.; Xiang, Y.; Newton, M. G.; Schinazi, R. F.; Cheng, Y.-C.; Chu, C. K. *J. Med. Chem.* **1997**, *40*, 3635–3644 and references cited therein.
 5. Yoshimura, Y.; Watanabe, M.; Satoh, H.; Ashida, N.; Ijichi, K.; Sakata, S.; Machida, H.; Matsuda, A. *J. Med. Chem.* **1997**, *40*, 2177–2183.
 6. a) Dyson, M. R.; Coe, P. L.; Walker, R. T. *J. Med. Chem.* **1991**, *34*, 2782–2786. b) Secrist, J. A.; Tiwari, K. N.; Riordan, J. M.; Montgomery, J. A. *J. Med. Chem.* **1991**, *34*, 2361–2366. c) Rahim, S. G.; Trivedi, N.; Bogunovic-Batchelor, M. V.; Hardy, G. W.; Mills, G.; Selway, J. W. T.; Snowden, W.; Littler, E.; Coe, P. L.; Basnak, I.; Whale, R. F.; Walker, R. T. *J. Med. Chem.* **1996**, *39*, 789–795. d) Van Draanen, N. A.; Freeman, G. A.; Short, S. A.; Harvey, R.; Jansen, R.; Szczech, G.; Koszalka, G. W. *J. Med. Chem.* **1996**, *39*, 538–542. e) Brånalt, J.; Kvarnström, I.; Niklasson, G.; Svensson, S. C. T.; Classon, B.; Samuelsson, B. *J. Org. Chem.*, **1994**, *59*, 1783–1788. f) Tber, B.; Fahmi, N.-E.; Ronco, G.; Villa, P.; Ewing, D. F.; Mackenzie, G. *Carbohydr. Res.*, **1995**, *267*, 203–215.
 7. a) Uenishi, J.; Takahashi, K.; Motoyama, M.; Akashi, H.; Sasaki, T. *Nucleosides Nucleotides* **1994**, *13*, 1347–1361. b) Young, R. J.; Shaw-Ponter, S.; Thomson, J. B.; Miller, J. A.; Cumming, J. G.; Pugh, A. W.; Rider, P. *Bioorg. Med. Chem. Lett.*, **1995**, *5*, 2599–2604.
 8. a) Yoshimura, Y.; Kitano, K.; Satoh, H.; Watanabe, M.; Miura, S.; Sakata, S.; Sasaki, T.; Matsuda, A. *J. Org. Chem.* **1996**, *61*, 822–823. b) Yoshimura, Y.; Kitano, K.; Yamada, K.; Satoh, H.; Watanabe, M.; Miura, S.; Sakata, S.; Sasaki, T.; Matsuda, A. *J. Org. Chem.* **1997**, *62*, 3140–3152.
 9. β -4: $[\alpha]_D^{25}$ -28.3° (c 0.13, H₂O) [cf. β -D-4: $[\alpha]_D^{25}$ $+24.7^\circ$ (c 0.18, H₂O)]; ¹H NMR (DMSO-*d*₆) δ 11.25 (1H, s, D₂O exchangeable), 7.93 (1H, d, *J* = 1.0 Hz), 6.07 (1H, d, *J* = 5.9 Hz), 5.69 (1H, d, *J* = 5.4 Hz, D₂O exchangeable), 5.40 (1H, d, *J* = 4.9 Hz, D₂O exchangeable), 5.21 (1H, t, *J* = 5.1 Hz, D₂O exchangeable), 4.00 (1H, q, *J* = 5.9 Hz), 3.94 (1H, q, *J* = 5.4 Hz), 3.75 (1H, dt, *J* = 4.9, 11.2 Hz), 3.66 (1H, dt, *J* = 5.9, 11.2 Hz), 3.13 (1H, q, *J* = 5.4 Hz), 1.77 (3H, s); FAB MS *m/z* 275 (M+H⁺). Anal. Calcd for C₁₀H₁₄N₂O₅S: C, 43.79; H, 5.14; N, 10.21. Found: C, 43.64; H, 5.31; N, 10.20. α -4: $[\alpha]_D^{25}$ -122.2° (c 0.13, H₂O) [cf. α -D-4: $[\alpha]_D^{25}$ $+116.8^\circ$ (c 0.15, H₂O)]; ¹H NMR (DMSO-*d*₆) δ 11.27 (1H, s, D₂O exchangeable), 7.84 (1H, s), 5.74 (1H, d, *J* = 7.8 Hz), 5.67 (1H, d, *J* = 5.7 Hz, D₂O exchangeable), 5.52 (1H, d, *J* = 4.9 Hz, D₂O exchangeable), 4.89 (1H, t, *J* = 5.1 Hz, D₂O exchangeable), 3.99 (1H, dt, *J* = 5.7, 7.8 Hz), 3.87–3.82 (1H, m), 3.60 (1H, dt, *J* = 4.9, 8.3 Hz), 3.52 (1H, dt, *J* = 3.4, 8.3 Hz), 3.40–3.35 (1H, m), 1.81 (3H, s); FAB MS *m/z* 275 (M+H⁺). Anal. Calcd for C₁₀H₁₄N₂O₅S: C, 43.79; H, 5.14; N, 10.21. Found: C, 43.50; H, 5.10; N, 9.82.
 10. β -5: $[\alpha]_D^{25}$ -74.0° (c 0.12, H₂O) [cf. β -D-5: *lit.* $[\alpha]_D^{25}$ $+72.8^\circ$; Ototani, N.; Whistler, R. L. *J. Med. Chem.* **1974**, *17*, 535–537.]; ¹H NMR (DMSO-*d*₆) δ 7.96 (1H, d, *J* = 7.8 Hz), 7.10, 7.01 (total 2H, brs, D₂O exchangeable), 6.33 (1H, d, *J* = 4.9 Hz), 5.69 (1H, d, *J* = 7.8 Hz), 5.56 (1H, d, *J* = 4.9 Hz, D₂O exchangeable), 5.35 (1H, d, *J* = 3.9 Hz, D₂O exchangeable), 5.05 (1H, t, *J* = 5.4 Hz, D₂O exchangeable), 3.98–3.92 (2H, m), 3.78 (1H, dt, *J* = 5.4, 11.2 Hz), 3.58 (1H, dt, *J* = 5.9, 11.2 Hz), 3.18–3.13 (1H, m); FAB MS *m/z* 260 (M+H⁺). Anal. Calcd for C₉H₁₃N₃O₄S•0.5H₂O: C, 40.29; H, 5.26; N, 15.66. Found: C, 40.22; H, 5.06; N, 15.38. α -5: $[\alpha]_D^{25}$ -136.3° (c 0.10, H₂O); ¹H NMR (DMSO-*d*₆) δ 7.89 (1H, d, *J* = 7.3 Hz), 7.14, 7.08 (total 2H, brs, D₂O exchangeable), 5.85 (1H, d, *J* = 7.3 Hz), 5.76 (1H, d, *J* = 7.3 Hz), 5.57 (1H, d, *J* = 5.9 Hz, D₂O exchangeable), 5.44 (1H, d, *J* = 4.9 Hz, D₂O exchangeable), 4.87 (1H, t, *J* = 5.4 Hz, D₂O exchangeable), 3.93 (1H, q, *J* = 6.8 Hz), 3.82 (1H, dt, *J* = 4.4, 10.7 Hz), 3.64 (1H, dt, *J* = 4.9, 7.3 Hz), 3.45 (1H, dt, *J* = 3.9, 7.8 Hz), 3.36 (1H, ddd, *J* = 5.9, 8.3, 10.7 Hz); FAB MS *m/z* 260 (M+H⁺). Anal. Calcd for C₉H₁₃N₃O₄S•0.25H₂O: C, 40.98; H, 5.16; N, 15.93. Found: C, 41.01; H, 5.04; N, 15.82.